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Biofilm and Trace Metals Removal from Waste Aqueous Effluent Using Moringa oleifera-Silver Nitrate Microparticles

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Authors' contributions

This work was carried out in collaboration between both authors. Authors ULN and ORR designed the study. Author ULN performed the statistical analysis and wrote the first draft of the manuscript. Author ORR wrote the protocol. Authors ULN and ORR managed the analyses of the study and literature searches. Both authors read and approved the final manuscript.

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ABSTRACT

The present research investigated the synthesis of a novel adsorbent by combining Moringa oleifera and silver nitrate $(AgNO₃)$. The chemical components of the pre-adsorbent and adsorbent composites were characterized using Fourier Transform Infra-red spectroscopy (FTIR). The equilibrium adsorption capacity (Qe) for Cu, Mn and Fe in DMS_{H2O} adsorbent was 0.04, 0.04 and 0.04 mg/g, respectively, while that for Cu, Mn and Fe for DMS_{C2H5OH} adsorbent was also 0.04, 0.04 and 0.04 mg/g, respectively. The results indicate that the adsorbent was effective in removing Cu more than Mn and Fe from the aqueous salt mixture. Furthermore, the results observed for the biofilm experiment revealed that the M . oleifera-AgNO₃ biocidal action was effective in deactivating oxidizable components in the wastewater system after 8 days. T-test of difference=0, at df=12, P<.05 for metal adsorption and t-test of difference=0.04, at df=8, P<.05 for COD removal indicate no

statistical significant difference in metals adsorbed and COD removed in the waste stream. The above findings have demonstrated that the synthesized M. oleifera-AgNO₃ adsorbent would be effective in cleaning-up waste streams contaminated with microbes and trace metals.

Keywords: Adsorption; contamination; environment; metals; plant; pollution; Moringa oleifera; wastewater.

1. INTRODUCTION

The need for high quality drinking water is increasing as non-polluted water sources are depleting due to effects of climate change and limited water sources. However, discharge criteria of wastewater are also becoming stricter since new legislations are coming on stream to prevent environmental pollution. Hence, water treatment has become very essential to meet the demand for portable water for consumption.

Trace metals are elements such as chromium (Cr), Cobalt (Co), Copper (Cu), Iron (Fe), Zinc (Zn) etc that normally occur at very low levels in the environment. But in higher amounts, many of the trace elements have been linked to instances of chronic liver and kidney injury [1]. Concentrations of trace metals in the environment increase when they are released from rocks and industrial wastes. These releases can occur through natural processes or through human activities. Human activities that release trace metals into the environment include; mining, smelting, burning of coal, and wastewater disposal [1].

Moringa oleifera in the family of Moringaceae, is a popular shrub grown mainly in tropic climates. Variety of uses of the plant has been reported severally [2]. Most importantly, the plant has been used as a water treatment natural bioactive compound extract and coagulant for turbid water [3]. Moringa oleifera plant has been studied extensively for its biosorption characteristics. Biosorption is a physiochemical process that occurs naturally in certain biomass which allows the biomass to passively concentrate and bind contaminants onto its cellular structure [4]. Biosorption provides an economical alternative for removing toxic heavy metals from domestic and industrial effluent.

According to Adelaja et al. [5], the adsorption of lead (II) ions (Pb^{2+}) onto ground M. oleifera pods was observed to be dependent on the pH and concentration of the Pb^{2+} aqueous solution as well adsorbent dosage and contact time of the adsorption process. However, the study concluded that M. oleifera pods were not effective as biosorbent for removal of Pb^{2+} ions from wastewater at pH 7.0. In a related study, M. oleifera seeds were investigated as biosorbent for removing metal ions from domestic effluent [6]. The authors noted that the plant has an efficient morphological profile for adsorbing ions (Cd, Pb, Cu) from waste streams due to the presence of interstices and protein component resulting in a high heterogeneity of the seed's matrix.

Trace metals such as Cd, has been successfully determined in alcohol fuel using M. oleifera seeds as biosorbent [2]. The plant can also be used as a natural solid adsorbent for removing silver iodide (AgI) in aqueous solutions [7]. Recent advances have come to light in the application of M. oleifera plants as biosorbents in investigations conducted by Sharma et al. [3] and Sharma et al. [8]. Both studies observed that the M. oleifera plant could be applied in waste aqueous systems to effectively remove Cd as well as in ternary biosorption studies to remove Cd(II), Cr(III) and Ni(II), respectively. However, the use of M. oleifera in combination with silver nitrate as a micro-adsorbent has not been reported.

Biofilm consists of organic and inorganic materials, ranging from decaying products in the wastewater [9]. Biofilm formation occurs when microorganisms attach themselves to subsurface structures in the water system. Biofilms develop quickly on iron than on plastic and polymer pipe surfaces. Iron pipes also encourage the composition of the microbial communities present in the biofilm [10]. In further studies, it was observed that biofilms in drinking water systems can serve as a reservoir for Helicobacter pylori (microorganism implicated in causing ulcers and cancer) [11]. Biofilms, while being a cause of concern for water utilities management, has been associated with the removal of some haloacetic acids (HAA) such as mono and dihalogenated species in drinking water distribution systems. HAA are commonly degraded by Afipia felis found in drinking water [12].

Several characteristics of the aqueous mixture such as pH, nutrients level, ionic strength, and temperature, have been noted to influence the rate of microbial attachment to a substratum, and hence increase biofilms proliferation [13]. Many researchers are of the opinion that a definite relationship exist between seasonal effects and biofilm formation in aqueous systems [14]. According to Fletcher [15], an increase in the concentration of Na⁺, Ca^{2+} , and Fe³⁺ ions in aqueous systems influenced the attachment of Pseudomonas fluorescens on glass surfaces; apparently, the ions reduced the repulsive forces between the negatively charged bacterial cells and the glass surfaces.

Silver (Ag) and silver ions (Ag^+) have been reported to have strong bacterial inhibition. Its biocidal effects when applied to pathogenic bacteria as disinfectant agents, is not in doubt [16,17].

The antimicrobial effect of Ag and $Ag⁺$ on different types of microorganisms has been investigated by several researchers [16,17]. To render the cell inactive, Ag and Ag⁺ binds to disulphide or sulfhydryl groups in the cell wall proteins and also to the deoxyribonucleic acid molecule in the cell. These binding actions disrupt metabolic processes in the cell and lead to cell death [18,19].

Silver has been applied as disinfectant in water filters against planktonic bacteria [20]. In combination with copper, Ag has been used effectively in water distribution systems against Legionella pneumophilia [21,22].

Silver and Ag nanoparticles (AgNPs) have general water disinfection anti-bacterial properties against a wide range of gram-negative and gram-positive bacteria, which include; Escherichia, Salmonella, Clostridium and Staphylococcus [23]. However, recent findings by Sun et al. [24] and Elechiquerra et al. [25] have suggested that AgNPs also has biocidal action against fungi such as Aspergillus niger, Candida albicans and viruses such as hepatitis B and the human immune deficiency virus (HIV-1). There are indications that high doses or repeated administration of Ag and Ag⁺ ions (approximately 10 g) can result in signs of mild poisoning considered to be of no toxicological significance [26].

Therefore, Moringa oleifera-AgNO₃ synergy, is an innovative combination when applied to treat water and wastewater.

2. MATERIALS AND METHODS

2.1 Adsorbent Preparation

Approximately 100 ml of a 0.1 mg/l silver nitrate $(AqNO₃)$ solution was measured into a 500 ml beaker, and 100 g of Moringa oleifera seeds, whose external kernel shells have been removed and the kernels previously washed and rinsed with deionized water were added to the $AgNO₃$ solution in the 500 ml beaker. About 10 ml solution of a 25% $\frac{v}{v}$ tetramethylsilane in ether was also measured and added to the M. oleifera- $A \cap A$ mixture. The mixture was allowed to stand for 48 h to allow the seeds adsorb the aqueous mixture. The M. oleifera seeds were recovered after this period and sun-dried for another 48 h and further placed in an oven (AEG 60 cm, Canberra, Australia) at 70ºC until constant weight.

Adsorbents of the M. oleifera seed were prepared by grinding the seeds manually to 5 mm. The ground seeds were divided into two parts. One part was washed in 10 ml of deionized H_2O , while the other part was washed in a 10% $\sqrt{7}$ solution of alcohol. Both washed seeds were dried in an oven at 50ºC for 1 h. Washing with these solvents introduces hydroxyl groups in the adsorbent, restrict particle agglomeration and increase adsorption capacity. The dried M. oleifera seeds were further labelled DMS_{H2O} and DMS_{C2H5OH} , respectively. About 1 g each of both DMS_{H2O} and DMS_{C2H5OH} were recovered to obtain the Infra-red spectra (Shidmazu FTIR, 8400S, Japan) of the preadsorbent materials. Adsorbents of DMS_{H2O} and DMS_{C2H5OH} were prepared by placing the entire quantity of both washed materials in a crucible and heat-treated in a furnace (Mini FurnaceTZ-1200C, Shanghai, China) at 200ºC for 1 h. The adsorbents were recovered from the furnace and cooled in a dessicator. About 1 g of the synthesized adsorbent each from DMS_{H2O} and DMS_{C2H5OH} were also withdrawn and used to obtain infra-red spectra of the synthesized adsorbent materials.

2.2 Equilibrium Adsorption Capacity Measurement (Qe)

Approximately 90 ml mixture consisting of a 30 ml solution of a 1 g/l each of the following salts $FeSO₄$, CuSO₄ and MnSO₄ was measured into a 200 ml beaker. The pH of the mixture was recorded using a pH meter (PHS 25, Shanghai Automation Instr. Corp., China) and adjusted to

 $pH=4$ using 20% ($\sqrt[w]{y}$) NaOH and 0.1 M HCl, added as appropriate. Exactly, 2 g of the synthesized DMS_{H2O} was weighed and added to the mixture in the 200 ml beaker. The adsorbentsalt solution mixture was stirred for 5 min and allowed to stand for 20 min. After this period about 5 ml of the mixture was filtered (Whatman No 42 filter paper) and used to determine the concentration of the metals (Fe, Cu, and Mn) in the mixture using a Multi-parameter photometer (H183200, HANNA Instr. UK). Three repetitions were made and the mean adsorption capacity (Q) of the adsorbent was estimated using the following equation:

$$
Q = \frac{(Co - Ce)(V)}{M}
$$
 (1)

Where

Co = Initial concentration of the salt used Ce = Concentration of the residual metal in

- the salt mixture after sorption
- $V =$ Volume of prepared salt mixture

 $M =$ Mass of adsorbent

The concentration of the residual metals in the salt mixture was determined intermittently at 10 min interval. A graph of Q versus time (t) was plotted, and the constant value of Q was referred as the equilibrium adsorption capacity (Qe).

This procedure was repeated for the synthesized DMS_{C2H5OH}.

2.3 Biofilm Experiment

The wastewater used for this research was obtained from a waste pipe channelling wastewater from the students Hostel A residence of the Federal University of Technology, Owerri, Nigeria.

A 1L plastic bottle was used to collect the wastewater and this was transported to the Project Laboratory of the Chemistry Department, Federal University of Technology, Owerri, Nigeria. About 100 ml each of the wastewater was poured into two separate 200 ml beakers, previously, washed and rinsed with distilled water and labelled BE 1 and BE 2.

About 10 ml of the wastewater in BE 1 was measured into a 50 ml conical flask. The pH of the wastewater was recorded using a pH meter (PHS 25, Shanghai Automation Instr. Corp., China) and adjusted to pH=8 using 2 ml of a 20%

 $(\sqrt[w]{v})$ NaOH, added as appropriate. Exactly, 3 ml of a 0.01 M potassium dichromate reagent and 7 ml of a10 M sulphuric acid reagent were separately measured and poured into the 50 ml conical flask.

The top of the flask was capped using a rubber cork. The mixture was stirred for 3 min and placed inside a chemical oxygen demand (COD) digester (2015D, Spectralab Intr. Ltd, India) at 110 $^{\circ}$ C for 30 min. After this period, the conical flask was withdrawn from the digester and cooled in a water bath (WBH14, GENLAB, UK) to room temperature. About 3 drops of ferroin indicator were added to the digestate in the conical flask and the content titrated against a 0.1 M ferrous ammonium sulphate (FAS) solution to a reddish brown color. Three repetitions were performed and the mean COD (mg/l) concentration was obtained. A control sample was processed as in above without adding the wastewater sample. The mean COD removed in the mixture was obtained using the following equation;

$$
COD = (X - Y) \times M \times 8000/ml
$$
 (2)

Where

- $X = mI$ of FAS solution for the control experiment
- $Y = mI$ of FAS solution for the sample
- $M =$ Molarity of FAS
- $8000 =$ Milli equivalent weight of oxygen x 1000 ml/l

Exactly, 2 g of the synthesized adsorbent DMS_{H2O} was added to BE 2. The mixture was stirred continuous for 10 min and the COD of the mixture was estimated after two days as described above for BE 1. This procedure was repeated for 10 days at interval of 2 days.

However, the procedure was also repeated using the synthesized DMS_{C2H5OH} adsorbent.

2.4 Statistical Analysis

Arithmetic mean and standard deviation were used to present data. The percentage relative error was used to estimate how closely measured metal concentration relates, while the t-test was also used to estimate significant difference in adsorbed trace metals concentrations between the DMS_{H2O} and DMS_{C2H5OH} synthesized adsorbents and DMS_{C2H5OH} synthesized adsorbents and concentrations of COD (mg/l) of wastewater at various times for synthesized adsorbent DMS_{H2O}

and DMS_{C2H5OH}. Data were analysed using Microsoft 2000 Excel (Version 9).

3. RESULTS AND DISCUSSION

Table 1 show the Infra-red spectra peak characteristics for DMS_{H2O} and DMS_{C2H5OH} pre-
adsorbent and synthesized adsorbent synthesized composites, respectively. The peaks around 3944 and 3813 $cm⁻¹$ represent medium and strong bonded OH groups, while that at 1750 cm⁻¹ is indicative of unsaturated ketone group for DMS_{H2O} pre-adsorbent composite. A slight shift of these peaks was observed around 3940-3755 cm^{-1} and 1748 cm^{-1} for the synthesized adsorbent composite. Nonetheless, the preadsorbent substance of DMS_{C2H5OH} showed peaks at 3906, 3817 and 1735, cm^{-1} , also representing medium and strong bonded OH groups and ketones. A slight shift of these peaks was also observed around 3975, 3770 and 1744 cm^{-1} for the synthesized DMS_{C2H5OH} material.

The peaks observed at 3334 cm^{-1} and 3381 cm^{-1} represents adsorption peaks for DMS_{H2O} and DMS_{C2H5OH} pre-adsorbent composites, respectively, while the peaks at 3357 cm^{-1} and 3384 cm⁻¹ also represents adsorption peaks for DMS_{H2O} and DMS_{C2H5OH} synthesized adsorbent materials, respectively. These adsorption peaks were assigned to N-H medium stretching primary amine. The presence of such functionality in the structure of the M. oleifera plant has been reported [27]. We can conclude that the shifts in peak values observed could be as a result of formation of chemical bond between functional groups and Ag⁺ ions during calcining. This line of reasoning is consistent with findings published by Bello et al. [28], Al-Ghouti et al. [29] and Bekci et al. [30].

The absorption peaks around $486-698$ cm⁻¹ and 528-613 $cm⁻¹$ represents adsorption peaks for DMS_{H2O} and DMS_{C2H5OH} synthesized adsorbent materials, respectively. These peaks are assigned to C-Ag⁺ ions bond formed during calcining. Formation of such sharp absorption C-Ag⁺ ion bond has been reported Sheibley et al. [31].

Table 2 shows concentrations (g/l) of residual metals in salt mixture at various times for synthesized adsorbent DMS_{H2O} and DMS_{C2H5OH} . Table 3 shows the effect of time (min) on the adsorption capacity (Q) of the synthesized adsorbent DMS_{H2O} and DMS_{C2H5OH}, respectively. The values obtained for the residual metals in the salt mixture raise an important issue with respect to pH dependence by adsorbents for metal adsorption. Given the values observed for the adsorption capacity in the present study, we conclude that the ability of the adsorbents to preferentially adsorbed metals may have been pH dependent. This finding is in agreement with recently published data by Salehzadeh (32), who observed that adsorption capacity of heavy metals by Xanthium pensylvanicum was pH dependent. It was observed that the equilibrium adsorption capacity (Qe) for Cu, Mn and Fe in DMSH2O adsorbent was 0.04, 0.04 and 0.04 mg/g, respectively, while that for Cu, Mn and Fe for DMS_{C2H5OH} adsorbent was 0.04, 0.04 and 0.04 mg/g, respectively. These results indicate that the adsorbent was able to remove, approximately, the same concentrations of Cu, Mn and Fe from the aqueous salt mixture and, hence, the Qe values of the three metals did not show much difference between their removal ratios. However, the results obtained also indicated that washing the M . oleifera-AgNO₃ substance with water or ethanol had no effect on the adsorption capacity of the synthesized adsorbent.

Table 4 shows values of concentrations of COD at various days for DMS_{H2O} and DMS_{C2H5OH} adsorbents, respectively. For DMS_{H2O} , the COD of the wastewater increased appreciably from day 1 to 4. The reason for this phenomenon could be due to the increase in biofilms growth as a result of the warm environment (increased temperature) in the laboratory and within the reacting system. Such growth condition of microbes has been reported [33]. However, it was observed that after day 4 the COD of the wastewater started decreasing due to the impact of the M . oleifera-AgNO₃ biocidal action.

It was observed for the DMS_{C2H5OH} adsorbent that the COD marginally increased from day 1 to day 2, due to reasons described as in above, but steadily decreased from day 4. As from day 8, the COD was zero (0) indicating that there was no bacteria activity in the experimental system, hence, no demand for oxygen, since no oxidizable components were present in the wastewater.

The percentage relative error for Cu, Mn and Fe for DMS_{H2O} and DMS_{C2H5OH} synthesized adsorbents are 3, 2 and 0.6% (DMS $_{H2O}$), and 0.7, 0.0 and 8% (DMS $_{C2H5OH}$), respectively. Values for Cu adsorption using DMS_{H2O} and DMS_{C2H5OH} adsorbents were used to test for significant

difference in metal adsorption by the synthesized adsorbents. T-test of difference=0, at df=12, P<.05 for metal adsorption and t-test of difference=0.04, at df=8, P<.05 for COD removal

indicate no statistical significant difference in metals adsorbed and COD removed in the waste stream.

Table 1. Infra-red spectra peak characteristics of DMS_{H2O} and DMS_{C2H5OH} pre-adsorbent and **synthesized adsorbent composite**

Table 2. Concentrations (g/l) of residual metals in salt mixture at various times for synthesized adsorbent DMSH2O and DMSC2H5OH

Table 3. Values of adsorption capacity (Q) at various times for synthesized adsorbent DMS_{H2O} **and DMSC2H5OH**

Table 4. Concentrations of COD (mg/l) of wastewater at various times for synthesized adsorbent DMSH2O and DMSC2H5OH

*COD value of raw wastewater for day $1 = 173$ mg/l

4. CONCLUSION

The present research had synthesized a novel adsorbent using M . oleifera-AgNO₃, for removing trace metals (Cu, Mn and Fe) from the aqueous salt mixtures as well as preventing proliferation of biofilms (microbes) in wastewater. The results obtained have shown that M. oleifera seeds are effective as composites adsorbent for removing metals from aqueous systems. The shift in the FTIR spectra obtained for the pre-adsorbent and adsorbent composites, and the values obtained for the equilibrium adsorption capacities (Qe) as well as COD, have demonstrated that the M. o leifera-AgNO₃ adsorbent was not just a simple mixture of both substances but that a synergic interaction existed between the M. oleifera and $AgNO₃$ compound which should have occurred during the synthesis of the composites. However, it was also observed in the present study that washing the synthesized adsorbent with either $H₂O$ or $C₂H₅OH$ didn't influence the adsorption capacity of either adsorbent.

Mn, Fe, and Cu were the elements of interest in the present study because, although not a danger to human health when present in water at concentrations within recommended guideline values, intentionally high presence or high intake of Mn, Fe and Cu may cause severe problems which include; biofouling in distribution networks, staining of laundry, nausea, liver and kidney damage, and even death [34,35].

DISCLAIMER

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Xiong X, Liu J, He W, Xia T, He P, Chen X, Yang K, Wang A. Dose effect relationship between drinking water fluoride levels and damage to liver and kidney functions in children. Environmental Research. 2007; 103:112-116.

- 2. Alves VN, Mosquetta R, Coelho NMM, Bianchin JN, Roux KCP, Martendal E, Carasek E. Determination of cadmium in alcohol fuel using Moringa oleifera seeds as a biosorbent in an on-line system coupled to FAAS. Talanta. 2010;80:1133- 1138.
- 3. Sharma P, Kumari P, Srivastava MM, Srivastava S. Removal of cadmium from aqueous system by shelled Moringa oleifera Lam. seed powder. Bioresource and Technology. 2006;97:299-305.
- 4. Ahalya N, Ramachandra TV, Kanamadi RD. Biosorption of heavy metals. Journal of Chemistry and Environment. 2003;7:71- 79.
- 5. Adelaja OA, Amoo IA, Aderibigbe AD. Biosorpton of lead (II) ions from aqueous solution using Moringa oleifera pods. Archives of Applied Science Research. 2003;3:50-60.
- 6. Araújo CST, Alves VN, Rezende HC, Almeida ILS, Assuncao RMN, Tarley CRT, Segatelli MG, Coelho NM. Characterization and use of Moringa oleifera seeds as biosorbent for removing metal ions from aqueous effluents. Water Science & Technology. 2010a;62:457-465.
- 7. Araujo CST, Melo EI, Alves VN, Coelho NMM. Moringa oleifera lam. seeds as a natural solid adsorbent for removal of AgI in aqueous solutions. Journal of the Brazilian Chemcial Soceity. 2010b;21: 1721-1732.
- 8. Sharma P, Kumari P, Srivastava MM, Srivastava S. Ternary biosorption studies of Cd(II), Cr(III) and Ni(II) on shelled Moringa oleifera seeds. Bioresource and Technology. 2007;98:474-477.
- 9. Characklis WG, Marshall KC. Biofilms 1st Ed. John Wiley & Sons, New York. 1990;341-394.
- 10. Haas CN, Meyer MA, Paller MS. The ecology of acid-fast organisms in water supply, treatment, and distribution system. Journal of the American Water Works Association. 1983;75:139-144.
- 11. Mackay WG, Gribbon LT, Barer MR, Reid DC. Biofilms in drinking water systems: A possible reservoir for Helicobacter pylori.

Journal of Applied Microbiology. 1999;85; 525-595.

- 12. Bayless W, Andrews RC. Biodegradation of six haloacetic acids in drinking water. Journal of Water and Health. 2008;16:15- 22.
- 13. Donlan RM. Biofilms: Microbial life on surfaces. Emerging Infections Diseases. 2002;8:881-889.
- 14. Fera P, Siebel MA, Characklis WG, Prieur D. Seasonal variations in bacterial colonization of stainless steel, aluminium, and polycarbonate surfaces in a seawater flow system. Biofouling. 1989;1:251- 261.
- 15. Fletcher M. Attachment of Pseudomonas fluorescens to glass and influence of electrolytes on bacterium-substratum separation distance. Journal of Bacteriology. 1988;170:2027-2030.
- 16. Cassels JM, Yahja MT, Gerba CP, Rose JB. Efficacy of a combined system of copper and silver and free chlorine for inactivation of Neagleria fowleri amoebas in water. Water Science and Technology. 1995;31:119-122.
- 17. Davis RI, Etris SF. The development and functions of silver in water purifications and disease control. Catalysis Today. 1997;36: 107-114.
- 18. Feng QL, Wu J, Chen GO, Cui FZ, Kim TN, Kim JO. A mechanistic study of the antibacterial effect of silver ions on Esherichia coli and Staphylococeus aureus. Journal of Biomedical Materials Research Part A. 2000;52:662-668.
- 19. Thurman RB, Gerba CP. The molecular mechanism of copper and silver ions disinfection of bacteria and viruses. Critical Reviews in Environmental Control. 1989;18:295-315.
- 20. Russell AD, Hugo WB. Antimicrobial activity and action of silver. Progress in Medicinal Chemistry. 1994;31:351-370.
- 21. Silvestry-Rodriguez N, Bright KR, Slack DC, Uhlmann DR, Gerba CP. Inactivation of Pseudomonas aeruginosa and Aeromonas hydrophilia by silver in tap water. Journal of Environmental Science and Health Part A. 2007;42:1-6.
- 22. Stout JE, Yu VL. Experiences of the first 16 hospitals using copper-silver ionization for Legionella control: Implications for the evaluation of other disinfection modalities.

Infection Control & Hospital Epidemiology. 2003;24:563–568.

- 23. Wijnhoven SWP, Peijenburg WJ, Herberts CA, Hagens WI, Oomen AG, Heugens E, Roszek B, Bisschops J, Gosens I, van de Meent D, Dekkers S, De Jong W, van Zijverden M, Sips A, Geertsma R. Nanosilver-a review of available data and knowledge gaps in human and environmental risk assessment. Nanotoxiciology. 2007;3:109-138.
- 24. Sun L, Singh A, Vig K, Pillai S, Singh S. Silver nanoparticles inhibits replication of respiratory syncytial virus. Journal of Biomedicine and Nanotechnology. 2008;4: 149-158.
- 25. Elechiquerra J, Burt J, Morones J, Camacho-Bragado A, Gao X, Lara H, Yacaman M. Interaction of silver nanoparticles with HIV-1. Journal of Nanobiotechnology' 2005.
- 26. Kim JS, Sung JH, Ji JH, Song KS, Lee JH, Kang CS, Yu IJ. In vivo genotoxicity of silver nanopaticles after 90-day silver nanoparticle inhalation exposure. Safety and Health at Work. 2011;2:34-38.
- 27. Amabye TG. Chemical compositions and nutritional value of Moringa oleifera available in the market of Mekelle. Journal of Food and Nutrition Sciences. 2015;3: 187-190.
- 28. Bello OS, Adegoke KA, Akinyunni OO. Preparation and characterization of novel adsorbent from Moringa oleifera leaf. Applied Water Science. 2015;2012b:1-11.
- 29. Al-Ghouti MA, Khraisheh MAM, Allen SJ, Ahmad MN. The removal of dyes from textile wastewater: A study of the physical characteristics and adsorption mechanisms of diatomaceous earth. Journal of Environmental Management. 2003;69:229-234.
- 30. Bekci Z, Seki Y, Cavas L. Removal of malachite green by using an invasive marine alga Caulerpa racemosa var. Cylindracea. Journal of Hazardous Material. 2009;161:1454-1460.
- 31. Sheibley DW, Fowler MH. Infrared spectra of various metal oxides in the region of 2 to 26 microns. NASA Technical Paper, TN – D – 3750. 1967;48-65.
- 32. Salehzadeh J. Removal of heavy metals Pb²⁺, Cu²⁺, Zn²⁺, Cd²⁺, Ni²⁺, Co²⁺ and Fe³⁺ from aqueous solutions by using Xanthium

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pensylvancum. Leonardo Journal of Sciences. 2013;23:97-104.

- 33. Eddleman H. Optimum temperature for growth of bacteria; 1998. Available: www.disket.com/indiana/b062 Accessed 25 September 2016
- 34. Casey J. Iron and manganese in water; occurrence, drinking water standards,

treatment options. Aquavarra Research Publications Water Engineering Papers. 2009;3:1-6.

35. Jantsch W, Kulig K, Rumack BH. Massive copper sulfate ingestion resulting in hepatotoxicity. Journal of Toxicology-Clinical Toxicology. 1985;102:585-588.

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